

## **Anti-C1q antibodies in systemic lupus erythematosus**

A-M Orbai, L Truedsson, G Sturfelt, O Nived, H Fang, GS Alarcón, C Gordon, JT Merrill, PR Fortin, IN Bruce, DA Isenberg, DJ Wallace, R Ramsey-Goldman, S-C Bae, JG Hanly, J Sanchez-Guerrero, AE Clarke, CB Aranow, S Manzi, MB Urowitz, DD Gladman, KC Kalunian, MI Costner, VP Werth, A Zoma, S Bernatsky, G Ruiz-Irastorza, MA Khamashta, S Jacobsen, JP Buyon, P Maddison, MA Dooley, RF Van Vollenhoven, E Ginzler, T Stoll, C Peschken, JL Jorizzo, JP Callen, SS Lim, BJ Fessler, M Inanc, DL Kamen, A Rahman, K Steinsson, AG Franks, Jr, L Sigler, S Hameed, N Pham, R Brey, MH Weisman, G McGwin, Jr, LS Magder and M Petri

*Lupus* published online 14 August 2014

DOI: 10.1177/0961203314547791

The online version of this article can be found at:  
<http://lup.sagepub.com/content/early/2014/09/02/0961203314547791>

---

Published by:



<http://www.sagepublications.com>

**Additional services and information for *Lupus* can be found at:**

**Email Alerts:** <http://lup.sagepub.com/cgi/alerts>

**Subscriptions:** <http://lup.sagepub.com/subscriptions>

**Reprints:** <http://www.sagepub.com/journalsReprints.nav>

**Permissions:** <http://www.sagepub.com/journalsPermissions.nav>

>> [OnlineFirst Version of Record - Sep 3, 2014](#)

[OnlineFirst Version of Record - Aug 14, 2014](#)

[What is This?](#)

**PAPER****Anti-C1q antibodies in systemic lupus erythematosus**

A-M Orbai<sup>1</sup>, L Truedsson<sup>2</sup>, G Sturfelt<sup>3</sup>, O Nived<sup>3</sup>, H Fang<sup>1</sup>, GS Alarcón<sup>4</sup>, C Gordon<sup>5</sup>, JT Merrill<sup>6</sup>, PR Fortin<sup>7</sup>, IN Bruce<sup>8,9</sup>, DA Isenberg<sup>10</sup>, DJ Wallace<sup>11</sup>, R Ramsey-Goldman<sup>12</sup>, S-C Bae<sup>13</sup>, JG Hanly<sup>14</sup>, J Sanchez-Guerrero<sup>15</sup>, AE Clarke<sup>16</sup>, CB Aranow<sup>17</sup>, S Manzi<sup>18</sup>, MB Urowitz<sup>19</sup>, DD Gladman<sup>19</sup>, KC Kalunian<sup>20</sup>, MI Costner<sup>21</sup>, VP Werth<sup>22</sup>, A Zoma<sup>23</sup>, S Bernatsky<sup>16</sup>, G Ruiz-Irastorza<sup>24</sup>, MA Khamashta<sup>25</sup>, S Jacobsen<sup>26</sup>, JP Buyon<sup>27</sup>, P Maddison<sup>28</sup>, MA Dooley<sup>29</sup>, RF Van Vollenhoven<sup>30</sup>, E Ginzler<sup>31</sup>, T Stoll<sup>32</sup>, C Peschken<sup>33</sup>, JL Jorizzo<sup>34</sup>, JP Callen<sup>35</sup>, SS Lim<sup>36</sup>, BJ Fessler<sup>4</sup>, M Inanc<sup>37</sup>, DL Kamen<sup>38</sup>, A Rahman<sup>9</sup>, K Steinsson<sup>39</sup>, AG Franks Jr<sup>27</sup>, L Sigler<sup>1</sup>, S Hameed<sup>1</sup>, N Pham<sup>1</sup>, R Brey<sup>40</sup>, MH Weisman<sup>41</sup>, G McGwin Jr<sup>4</sup>, LS Magder<sup>42</sup> and M Petri<sup>1</sup>

<sup>1</sup>Division of Rheumatology, Johns Hopkins University School of Medicine, Baltimore, MD, USA; <sup>2</sup>Department of Laboratory Medicine, Section of Microbiology, Immunology and Glycobiology, Lund University, Lund, Sweden; <sup>3</sup>Department of Rheumatology, Skåne University Hospital, Lund, Sweden; <sup>4</sup>Department of Medicine, Division of Clinical Immunology and Rheumatology, University of Alabama at Birmingham, Birmingham, AL, USA; <sup>5</sup>Rheumatology Research Group, School of Immunity and Infection, College of Medical and Dental Sciences University of Birmingham, Birmingham, UK; <sup>6</sup>Department of Clinical Pharmacology, Oklahoma Medical Research Foundation, Oklahoma City, OK, USA; <sup>7</sup>Division of Rheumatology, Department of Medicine, Centre Hospitalier Universitaire (CHU) de Québec Axe Maladies Infectieuses et Immunitaires, CRCHU de Québec, Université Laval, Québec City, Québec, Canada; <sup>8</sup>Arthritis Research UK Centre for Epidemiology, Centre for Musculoskeletal Research, Institute of Inflammation and Repair, Manchester Academic Health Sciences Centre, The University of Manchester, Manchester, UK; <sup>9</sup>NIHR Manchester Musculoskeletal Biomedical Research Unit, Central Manchester University Hospitals NHS Foundation Trust, Manchester Academic Health Science Centre, UK; <sup>10</sup>Centre for Rheumatology, Research Division of Medicine, London, UK; <sup>11</sup>Cedars-Sinai Medical Center, David Geffen School of Medicine, University of California, Los Angeles, CA, USA; <sup>12</sup>Northwestern University, Feinberg School of Medicine, Chicago, IL, USA; <sup>13</sup>Department of Rheumatology, Hanyang University Hospital for Rheumatic Diseases, Seoul, Korea; <sup>14</sup>Division of Rheumatology, Departments of Medicine and Pathology Capital Health and Dalhousie University, Halifax, Nova Scotia, Canada; <sup>15</sup>Mount Sinai Hospital and University Health Network, Toronto, Ontario, Canada; <sup>16</sup>Divisions of Clinical Epidemiology and Rheumatology, McGill University Health Centre, Montreal, Quebec, Canada; <sup>17</sup>Feinstein Institute for Medical Research, Manhasset, NY, USA; <sup>18</sup>Department of Medicine, Division of Rheumatology, Allegheny Singer Research Institute, Allegheny General Hospital, Pittsburgh, PA, USA; <sup>19</sup>Toronto Western Hospital Toronto, Ontario, Canada; <sup>20</sup>Division of Rheumatology, Allergy and Immunology, UCSD School of Medicine, La Jolla, CA, USA; <sup>21</sup>North Dallas Dermatology Associates, Dallas, TX, USA; <sup>22</sup>Philadelphia VA Medical Center and University of Pennsylvania, Philadelphia, PA, USA; <sup>23</sup>Lanarkshire Centre for Rheumatology and Hairmyres Hospital, East Kilbride, UK; <sup>24</sup>Autoimmune Diseases Research Unit, Hospital Universitario Cruces Universidad del País Vasco, Barakaldo, Spain; <sup>25</sup>Rayne Institute and St Thomas' Hospital London, UK; <sup>26</sup>Rigshospitalet, Copenhagen University Hospital, Copenhagen, Denmark; <sup>27</sup>New York University, New York, NY, USA; <sup>28</sup>Ysbyty Gwynedd, Bangor, UK; <sup>29</sup>University of North Carolina, Chapel Hill, NC, USA; <sup>30</sup>Karolinska University Hospital, Stockholm, Sweden; <sup>31</sup>State University of New York, Downstate Medical Center, Brooklyn, NY, USA; <sup>32</sup>Kantonsspital Schaffhausen, Schaffhausen, Switzerland; <sup>33</sup>University of Manitoba Winnipeg, Manitoba, Canada; <sup>34</sup>Wake Forest University, Winston-Salem, NC, USA; <sup>35</sup>University of Louisville, Louisville, KY, USA; <sup>36</sup>Emory University, Atlanta, GA, USA; <sup>37</sup>Division of Rheumatology, Department of Internal Medicine, Istanbul Medical Faculty, Istanbul University, Istanbul, Turkey; <sup>38</sup>Medical University of South Carolina, Charleston, SC, USA; <sup>39</sup>Landspítali University Hospital, Reykjavik, Iceland; <sup>40</sup>University of Texas Health Science Center, San Antonio, TX, USA; <sup>41</sup>Cedars-Sinai Medical Center, Los Angeles, CA, USA; and <sup>42</sup>Department of Epidemiology and Public Health, University of Maryland, Baltimore, MD, USA

**Objective:** Anti-C1q has been associated with systemic lupus erythematosus (SLE) and lupus nephritis in previous studies. We studied anti-C1q specificity for SLE (vs rheumatic disease controls) and the association with SLE manifestations in an international multicenter study. **Methods:** Information and blood samples were obtained in a cross-sectional study from patients with SLE ( $n = 308$ ) and other rheumatologic diseases ( $n = 389$ ) from 25 clinical sites (84% female, 68% Caucasian, 17% African descent, 8% Asian, 7% other). IgG anti-C1q against the collagen-like region was measured by ELISA. **Results:** Prevalence of anti-C1q was 28% (86/308) in patients with SLE and 13% (49/389) in controls (OR = 2.7, 95% CI: 1.8–4,  $p < 0.001$ ). Anti-C1q was associated with proteinuria (OR = 3.0, 95% CI: 1.7–5.1,  $p < 0.001$ ), red cell casts (OR = 2.6, 95% CI: 1.2–5.4,  $p = 0.015$ ), anti-dsDNA (OR = 3.4, 95% CI: 1.9–6.1,  $p < 0.001$ ) and anti-Smith (OR = 2.8, 95% CI: 1.5–5.0,  $p = 0.01$ ). Anti-C1q was independently associated with renal involvement after adjustment for demographics, ANA, anti-dsDNA and low complement (OR = 2.3, 95% CI: 1.3–4.2,  $p < 0.01$ ). Simultaneously positive anti-C1q, anti-dsDNA and low complement was strongly associated with renal involvement (OR = 14.9, 95% CI: 5.8–38.4,  $p < 0.01$ ). **Conclusions:** Anti-C1q was more common in

Correspondence to: Michelle Petri, Division of Rheumatology Johns Hopkins University School of Medicine 1830 E. Monument St., Suite 7500, Baltimore, MD, USA.

Email: MPetri@jhmi.edu

Received 21 December 2013; accepted 28 July 2014

patients with SLE and those of Asian race/ethnicity. We confirmed a significant association of anti-C1q with renal involvement, independent of demographics and other serologies. Anti-C1q in combination with anti-dsDNA and low complement was the strongest serological association with renal involvement. These data support the usefulness of anti-C1q in SLE, especially in lupus nephritis. *Lupus* (2014) **0**, 1–8.

**Key words:** Anti-dsDNA antibodies; renal lupus; systemic lupus erythematosus

## Introduction

Complement plays a major role in the pathogenesis of systemic lupus erythematosus (SLE) and lupus nephritis. Genetic deficiencies in the early complement components are associated with SLE.<sup>1,2</sup> The strongest association is seen in patients with homozygous C1q deficiency, of whom 88% developed SLE and 30% glomerulonephritis, respectively.<sup>3</sup> In vitro, physiologic concentrations of C1q inhibit interferon alpha production by plasmacytoid dendritic cells stimulated with nucleic acid-containing immune complexes,<sup>4</sup> suggesting a regulatory effect of C1q in response to and clearance of immune complexes. In patients with SLE, levels of C1q were reduced in glomerulonephritis flares.<sup>5</sup> In patients with lupus nephritis, the presence of anti-C1q at the time of renal biopsy was associated with worse renal outcome, by the American College of Rheumatology (ACR) renal response criteria,<sup>6</sup> and with renal tubulointerstitial changes.<sup>7</sup> Acquired antibodies against the collagen-like region of C1q (anti-C1qCLR) were present in the glomerular basement membrane of patients with proliferative lupus nephritis at concentrations more than 50-fold higher per unit immunoglobulin (Ig)G than in the patients' serum, suggesting a role in the pathogenesis of lupus nephritis.<sup>8</sup> C1q were aggregated within IgG in renal subendothelial deposits in active proliferative lupus nephritis as seen on immunogold electron microscopy, further supporting a pathogenic role of anti-C1q.<sup>9</sup> Patients with active lupus nephritis had a higher prevalence of anti-C1q than those without lupus nephritis, 74% vs 32% ( $p < 0.0001$ ).<sup>10</sup> Anti-C1q increased within six months prior to renal involvement in 50% of patients with SLE<sup>11</sup> and was associated with the proliferative form of glomerulonephritis.<sup>12–14</sup> In another study, an increase in anti-C1q level preceded renal flare by 2.3 months and was more specific for renal flare than increases in anti-double-stranded DNA (anti-dsDNA) level.<sup>15</sup> Anti-C1q concentration correlated with activity on the modified Safety of Estrogen in Lupus: National Assessment-Systemic Lupus Erythematosus Disease Activity Index (SELENA-SLEDAI) and the Systemic Lupus International Collaborating

Clinics (SLICC) Renal Activity Score.<sup>16</sup> With immunosuppressive treatment for membranoproliferative lupus nephritis with either cyclophosphamide or azathioprine, anti-C1q disappeared by week 12 and remained undetectable throughout one year of follow-up.<sup>17</sup> As detailed above, evidence suggests that anti-C1q is associated not only with lupus nephritis, but also with lupus nephritis flares and response to treatment. Therefore anti-C1q might be a candidate for predicting lupus nephritis and monitoring treatment in clinical practice. The purpose of this study was to characterize, in a multinational patient population, the prevalence and clinical associations of anti-C1q in patients with SLE and other rheumatic diseases and to define the association of anti-C1q with renal involvement in patients with SLE.

## Patients and methods

### Patients

We studied anti-C1q specificity for SLE (vs rheumatic disease controls) and its association with SLE manifestations in an international, multicenter, cross-sectional sample of patients with SLE and other rheumatic diseases, assembled to derive the SLICC classification criteria for SLE.<sup>18</sup>

### Laboratory methods

Anti-C1q determination was performed at the laboratory of Lennart Truedsson, MD, PhD (Department of Laboratory Medicine, Section of Microbiology, Immunology and Glycobiology, Lund University, Lund, Sweden). An enzyme-linked immunosorbent assay (ELISA) with purified collagenous C1q fragments in the solid phase was used for detection of anti-C1q IgG in all serum samples obtained at the beginning of the study. The assay was previously described,<sup>19</sup> and it is well documented that autoantibodies against C1q in SLE target the collagenous portion of the molecule.<sup>20,21</sup> Use of purified C1q collagenous fragments as antigen in the ELISA prevented nonspecific interactions. The reference interval was defined as  $< 16$  AU/l based on analysis of anti-C1q IgG in 96 healthy blood donors.<sup>22</sup>

Laboratory determinations were performed at the Rheumatology Diagnostic Laboratory (Los Angeles, CA) for anti-dsDNA by ELISA, *Crithidia* assay and Farr assay, and for anti-Smith antibody and complement C3 and C4 levels. Another set of blood samples was tested for anti-phospholipid antibodies (lupus anticoagulant, and ELISA for IgG, IgM and IgA isotypes of anticardiolipin antibodies and anti- $\beta$ 2-glycoprotein I antibodies) at the laboratory of Joan Merrill, MD (Oklahoma Medical Research Foundation, USA).

### Statistical methods

Statistical analyses were carried out using SAS<sup>®</sup> (SAS<sup>®</sup> 9.2, SAS Institute Inc, Cary, NC, USA) and Stata statistical software (Stata 12, StataCorp LP, College Station, TX, USA). Patients with SLE and controls with rheumatic disease were compared with respect to demographic characteristics, clinical manifestations, and serologic results using Chi-square tests; *p* values for Chi-square tests were adjusted for age and ethnicity as specified in the tables. A *p* value  $\leq 0.05$  was considered statistically significant. In patients with SLE, we calculated odds ratios of renal involvement by the SLICC classification criteria (urine protein to creatinine ratio or 24-hour urine protein representing 500 mg/24 hours or red blood cell casts),<sup>23</sup> using multiple logistic regression on demographic and serologic characteristics. The first model adjusted for demographics (age, ethnicity, gender) and individual antibodies (antinuclear antibodies (ANA), anti-dsDNA, low complement C3 and/or C4 and anti-C1q). The second model adjusted for demographics and serologic patterns for anti-C1q, anti-dsDNA and low complement C3 and/or C4.

The study was approved by institutional review boards at all institutions involved, and all participants provided written informed consent.

## Results

Clinical information and blood samples were obtained from 308 patients with SLE (mean age (SD) 34 (13) years, 89% female, 63% Caucasian, 22% African descent, 12% Asian, 3% other) and 389 patients with other rheumatologic diseases (mean age (SD) 43 (15) years, 80% female, 73% Caucasian, 13% African descent, 5% Asian, 9% other) from 25 clinical sites. SLICC renal involvement was present in 33% of patients with SLE and 4% of controls. Of 308 patients with SLE, 72 (23%)

had biopsy-confirmed lupus nephritis (and none of the controls).

### Anti-C1q prevalence by diagnosis

The prevalence of anti-C1q was 28% (86/308) in patients with SLE and 13% (49/389) in controls with other rheumatologic disorders (OR = 2.7, 95% confidence interval (CI) 1.8–4.0, *p* < 0.001). The frequency of anti-C1q in rheumatic disease controls was: 26% in scleroderma, 19% in rheumatoid arthritis, 15% in undifferentiated connective tissue disease, 15% in chronic cutaneous lupus, 14% in Sjögren's syndrome, 8% in fibromyalgia, 7% in antiphospholipid antibody syndrome, 6% in dermatomyositis, and 5% in vasculitis.

### Anti-C1q and demographic characteristics in patients with SLE

Anti-C1q was more common in Asians (*n* = 37, 40.5%) than in Caucasians (*n* = 192, 27.6%) or patients of African descent (*n* = 69, 21.7%), but these differences were not statistically significant. Anti-C1q was more common in younger individuals with SLE, using an age cutoff of 30 years (35.5% vs 23%, *p* = 0.02) (Table 1).

### Anti-C1q and clinical SLE manifestations

Sensitivity of anti-C1q for a classification of SLE was 28% and specificity was 87%. In an age-adjusted analysis we assessed the clinical features of SLE associated with anti-C1q antibodies. Patients with anti-C1q were significantly more likely to have proteinuria (OR = 3.0, 95% CI 1.7–5.1, *p* < 0.001) and urinary red cell casts (OR = 2.6, 95% CI 1.2–5.4, *p* = 0.015). There was a trend towards an association with psychosis (OR = 9.5, 95% CI 0.9–98.5, *p* = 0.06). No significant associations were seen with arthritis, cutaneous lupus or hematologic manifestations (Table 2).

### Anti-C1q and serologic SLE manifestations

In patients with SLE positive for anti-C1q (compared to patients negative for anti-C1q), there were positive associations with anti-dsDNA (OR = 3.4, 95% CI 1.9–6.1, *p* < 0.001) and anti-Smith (OR = 2.8, 95% CI 1.5–5.0, *p* = 0.01) and no association with antiphospholipid antibodies after adjustment for age (Table 2).

### Anti-C1q and lupus nephritis

Sensitivity of anti-C1q for SLE renal involvement was 41% and specificity was 85%. Anti-C1q

**Table 1** Association between demographic characteristics and anti-C1q in SLE: Percentage of patients with anti-C1q, by demographic variables

Demographics	Percentage for anti-C1q	p value
Ethnicity		
African descent	21.7	0.15
Caucasian	27.6	
Asian	40.5	
Other	30.0	
Gender		
Female	26.9	0.25
Male	36.4	
Age (years)		
≤30	35.5	0.01
>30	23.0	

SLE: systemic lupus erythematosus.

**Table 2** Association between ACR criteria and anti-C1q in SLE: Percentage of patients with various clinical conditions, by anti-C1q status

ACR criteria	Anti-C1q positive (%)	Anti-C1q negative (%)	p value	Odds ratio (95% CI)	Adjusted p value for age
Malar rash	47.7	46.9	0.90	0.9 (0.5, 1.5)	0.69
Discoid rash	19.8	19.4	0.94	1.1 (0.6, 2.1)	0.71
Photosensitivity	53.5	53.2	0.96	1.0 (0.6, 1.7)	1.00
Oral ulcers	38.4	46.4	0.20	0.7 (0.4, 1.1)	0.14
Arthritis	64.0	65.8	0.76	0.9 (0.5, 1.5)	0.70
Serositis	37.2	34.7	0.68	1.1 (0.6, 1.8)	0.84
Pleurisy	31.4	28.4	0.60	1.1 (0.6, 1.9)	0.74
Pericarditis	14.0	12.2	0.67	1.2 (0.6, 2.5)	0.66
Proteinuria	50.0	22.5	<0.01	3.0 (1.7, 5.1)	<b>&lt;0.01</b>
Red cell casts	18.6	7.2	<0.01	2.6 (1.2, 5.4)	<b>0.02</b>
Seizure	5.8	4.1	0.51	1.2 (0.4, 3.8)	0.72
Psychosis	3.5	0.5	0.04	9.5 (0.9, 98.5)	0.06
Hematologic	64.0	58.1	0.35	1.2 (0.7, 2.0)	0.49
Leukopenia	40.7	35.1	0.36	1.2 (0.7, 2.0)	0.48
Lymphopenia	38.4	36.5	0.76	1.1 (0.7, 1.8)	0.73
Thrombocytopenia	15.1	12.2	0.49	1.1 (0.5, 2.2)	0.86
Anti-dsDNA	77.9	47.8	<0.01	3.4 (1.9, 6.1)	<b>&lt;0.01</b>
Anti-Smith	33.7	14.4	<0.01	2.8 (1.5, 5.0)	<b>0.01</b>
Antiphospholipid	57.0	54.5	0.70	1.1 (0.7, 1.8)	0.70

ACR: American College of Rheumatology; SLE: systemic lupus erythematosus; Anti-dsDNA: anti-double-stranded DNA; CI: confidence interval. Bold values represent  $p < 0.05$ .

prevalence in patients with SLE with, vs without ACR renal disorder (persistent proteinuria  $>0.5$  g/24 hour or proteinuria  $>3+$ , or red blood cell casts) was 45.5% compared to 19.3%, respectively (OR = 3.2, 95% CI 1.8–5.6,  $p < 0.001$ ). Additional serologic associations observed for ACR renal disorder were with anti-dsDNA (OR = 4.7, 95% CI

2.5–8.6,  $p < 0.001$ ), low complement (OR = 2.8, 95% CI 1.5–4.9,  $p = 0.001$ ) and anti-Smith (OR = 1.9, 95% CI 1.1–3.6,  $p = 0.03$ ), after adjustment for age and ethnicity (Table 3).

The first logistic regression model was applied to all patients with SLE ( $n = 308$ ) to estimate the independent contribution of demographic characteristics and serologies to odds of SLICC renal involvement ( $n = 101$ ). Odds of SLICC renal involvement were two times lower in patients above age 30 than below age 30 (OR = 0.4, 95% CI 0.3–0.8,  $p = 0.005$ ) and, independently of age, three times lower in Caucasians compared to African Americans (OR = 0.3, 95% CI 0.1–0.6,  $p < 0.001$ ), after adjustment for gender and serologies (Table 4).

Odds of SLICC renal involvement in the presence of anti-dsDNA were four times higher than in the absence of anti-dsDNA, after adjustment for age, ethnicity, gender and serologies (OR = 4.1, 95% CI 2.1–7.9,  $p < 0.001$ ). In the same model, for anti-C1q positive, odds of SLICC renal involvement were independently 2.3 times higher than in the absence of anti-C1q (OR = 2.3, 95% CI 1.3–4.2,  $p = 0.007$ ) (Table 4). Low complement C3 and/or C4, compared to normal, was associated with double the odds of SLICC renal involvement, a finding that was not statistically significant after adjustment for anti-dsDNA and anti-C1q (OR = 1.9, 95% CI 1.0–3.6,  $p = 0.06$ ).

The second logistic regression model estimated odds of SLICC renal involvement using possible combinations of serology results for anti-dsDNA, anti-C1q and low complement and adjusted for age, ethnicity and gender. By patterns of positivity for anti-C1q, anti-dsDNA and low complement, odds of SLICC renal involvement were 15 times higher for patients with all three serologies positive compared to all negative (OR = 14.9, 95% CI 5.8–38.4,  $p < 0.001$ ). In the same model, combinations of simultaneously positive anti-dsDNA and low complement, and simultaneously positive anti-dsDNA and anti-C1q were associated with five times and six times increase in odds of SLICC renal involvement than all three serologies negative, respectively (OR = 5.2 95% CI 2.1–13.1,  $p < 0.001$  and OR = 5.7 95% CI 1.2–28.3,  $p = 0.03$ ) (Table 5).

## Discussion

Anti-C1q has been associated with SLE and SLE nephritis in previous studies.<sup>10,12,14,24–28</sup> We confirmed this association in the SLICC international

**Table 3** Association with renal involvement: Percentage of patients with SLE serologies among those with and without ACR lupus nephritis

Variable	Renal involvement (%)	No renal involvement (%)	p value	Odds ratio (95% CI)	Adjusted p value for age and ethnicity
Anti-C1q	45.5	19.3	<0.01	3.2 (1.8, 5.6)	<0.01
Anti-dsDNA	80.2	44.4	<0.01	4.7 (2.5, 8.6)	<0.01
Anti-Smith	29.7	15.0	<0.01	1.9 (1.1, 3.6)	<b>0.03</b>
Low complement	78.2	50.2	<0.01	2.8 (1.5, 4.9)	<0.01

ACR: American College of Rheumatology; SLE: systemic lupus erythematosus; Anti-dsDNA: anti-double-stranded DNA; CI: confidence interval. Bold values represent  $p < 0.05$ .

**Table 4** Odds ratios (OR) of SLICC renal involvement<sup>a</sup> in patients with SLE ( $N = 308$ ) by individual antibody status, adjusted for demographic and serologic characteristics

Covariates	SLICC renal (OR)	95% CI	p value
<i>Age (years)</i>			
≤30 (ref.) <sup>b</sup>	1.00	0.25–0.78	<0.01
>30	0.44		
<i>Ethnicity</i>			
African American (ref.)	1.00	0.14–0.60	<0.01
White	0.28	0.16–1.08	0.07
Asian	0.42	0.09–1.71	0.22
Hispanic/Latino	0.40		
<i>Gender</i>			
Male (ref.)	1.00	0.19–1.14	0.09
Female	0.46		
ANA	0.22	0.05–1.01	0.05
Anti-dsDNA	4.05	2.10–7.90	<0.01
Low complement	1.87	0.98–3.59	0.06
Anti-C1q	2.30	1.26–4.19	<0.01

SLICC: Systemic Lupus Collaborating Clinics; SLE: systemic lupus erythematosus; Anti-dsDNA: anti-double-stranded DNA; CI: confidence interval.

<sup>a</sup>SLICC renal involvement is defined as urine protein-to-creatinine ratio (or 24-hour urine protein) representing 500 mg/24 hours or red blood cell casts; estimates from multivariable logistic regression model, constant term 3.44, 95% CI 0.51–23.07,  $p$  value 0.2).

<sup>b</sup>ref.: the reference group for each category. Bold values represent  $p < 0.05$ .

patient population, of whom we studied 308 patients with SLE and 389 controls with other rheumatologic diseases. We also showed, for the first time, the association of anti-C1q with lupus renal involvement by SLICC classification criteria.

The presence of anti-C1q antibodies in other autoimmune diseases, as we have found, and even in healthy individuals (4% to 6.4%), has been previously reported.<sup>21,29</sup> Patients with scleroderma were anti-C1q positive in a higher proportion in our study, 26%, than observed in other studies, 5.5%.<sup>24</sup> None of these patients had renal involvement. Patients with rheumatoid arthritis were anti-C1q positive in a higher proportion in our study,

19%, than observed in other studies, 5%.<sup>30</sup> However, a review by Seelen et al. reported anti-C1q prevalence of 77% in rheumatoid vasculitis.<sup>31</sup> Patients with vasculitis were less often positive in the population we studied, 5%, vs 12%–35% in other studies.<sup>29</sup> We did not collect information on type of vasculitis and anti-neutrophil cytoplasmic antibody (ANCA) status. Anti-C1q has not been previously described in dermatomyositis in which we found a prevalence of anti-C1q of 6% (based on 55 patients). Anti-C1q was more common in Asians (40.5%) than in Caucasians (27.6%) and patients of African descent (21.7%), but these differences were not statistically significant, consistent with previous studies.<sup>32</sup> We found that younger individuals with SLE were more likely to be anti-C1q positive than older individuals, using an age cutoff of 30 years. Siegert et al., similarly, found a higher prevalence of IgG anti-C1q antibodies in younger individuals with SLE compared to random selected controls (highest titer and highest prevalence below age 30); in patients with SLE anti-C1q prevalence decreased with age while in random controls the opposite was true.<sup>33</sup> Anti-C1q antibody prevalence in patients with SLE with ACR renal involvement was 45.5% in our study. Braun et al. found a prevalence of 61.7% in biopsy-proven lupus nephritis cases<sup>29</sup> and Wener et al., 48%.<sup>21</sup> The strongest clinical association we observed for anti-C1q was with proteinuria, consistent with published data.<sup>12,14,16,27,34</sup> Our study was undertaken in patients with SLE from a multi-center, multiethnic patient population and a similar number of patients with other rheumatic diseases (controls), in which complete clinical, serologic and candidate criteria variables were assessed for the purpose of deriving SLE classification rules. We did not have flare data, treatment data, or repeat anti-C1q antibody levels because of the cross-sectional nature of this study. Therefore, any temporal relationship of anti-C1q antibody levels to flares of lupus nephritis or change in treatment could not be

**Table 5** Odds ratios (OR) of SLICC renal involvement<sup>a</sup> in patients with SLE (*N* = 308) by antibody patterns, adjusted for demographics (age, ethnicity and gender)

<i>Serologic patterns</i>			<i>N</i> (308)	<i>SLICC renal (OR*)</i>	<i>95% CI</i>	<i>p value</i>
<i>Anti-C1q</i>	<i>Anti-dsDNA</i>	<i>Low complement</i>				
Negative (ref.) <sup>b</sup>	Negative	Negative	75	1.00		
Negative	Negative	Positive	41	1.83	0.62–5.34	0.27
Negative	Positive	Negative	35	2.46	0.79–7.61	0.12
Negative	Positive	Positive	71	5.23	2.10–13.05	<b>&lt;0.01</b>
Positive	Negative	Negative	6	4.06	0.60–27.30	0.15
Positive	Negative	Positive	13	0.66	0.07–6.11	0.72
Positive	Positive	Negative	9	5.74	1.16–28.29	<b>0.03</b>
Positive	Positive	Positive	58	14.89	5.77–38.44	<b>&lt;0.01</b>

SLICC: Systemic Lupus Collaborating Clinics; SLE: systemic lupus erythematosus.

<sup>a</sup>SLICC renal involvement is defined as urine protein-to-creatinine ratio (or 24-hour urine protein) representing 500 mg/24 hours or red blood cell casts; estimates of odds ratios (OR) are from multivariable logistic regression model, constant term 0.95, 95% CI 0.26–3.49, *p* value 0.94).

<sup>b</sup>ref.: the reference group for each category. Bold values represent *p* < 0.05.

assessed. Others noted that anti-C1q antibody levels increased prior to flares of lupus nephritis and disappeared with immunosuppressive treatment.<sup>11,15,26</sup> Moroni *et al.* showed an association with active lupus nephritis for anti-C1q and low complement.<sup>14</sup> Yang *et al.* showed concomitant presence of anti-C1q and anti-dsDNA was associated with higher lupus nephritis activity and poor renal outcome compared to only one or none of these antibodies.<sup>28</sup> Anti-C1q in our study had the highest prevalence in patients with SLE with ACR renal involvement and was strongly associated with anti-dsDNA and low complement. It was the second highest antibody associated with a diagnosis of ACR renal involvement, after anti-dsDNA. By the SLICC classification criteria, age above 30 years and being Caucasian were protective from SLICC renal involvement: These characteristics were independently associated with decreased odds of renal involvement in patients with SLE by two and three times, respectively, which is consistent with the literature on the subject. Independently of each other, anti-dsDNA (vs negative) was associated with four times higher odds of SLICC renal involvement and anti-C1q (vs negative) was associated with two times higher odds of SLICC renal involvement, after adjustment for age, ethnicity, gender and low complement. In patients with SLE, odds of SLICC renal involvement were highest in the presence of simultaneously positive anti-dsDNA, anti-C1q and low complement (15 times higher than all negative). Increases in odds of SLICC renal involvement with concomitantly positive anti-dsDNA and low complement were similar to concomitantly positive anti-dsDNA and anti-C1q. As seen in the logistic

regression models, the three serologies (anti-C1q, anti-dsDNA, low complement) had a multiplicative relationship in increasing the odds of SLICC renal involvement after adjustment for demographics.

Many studies of anti-C1q antibodies are performed with methods using whole C1q molecules as antigen and a buffer with high ionic strength to prevent nonspecific interaction between the globular heads of C1q and antibodies. In this study the purified collagenous fragment was used as antigen in the ELISA and the nonspecific interactions were thereby avoided.<sup>19</sup> Comparisons between the method used here and the high-salt buffer method in 100 patients with high and low disease activity have given very similar results (Truedsson *et al.* 2014, personal communication). The reason anti-C1q was eliminated, at the end of the derivation phase of the SLICC classification criteria for SLE, was mainly because of a lack of a high-quality, standardized, less laborious assay. As new laboratory techniques develop and further uses of anti-C1q determinations become important for clinical care and disease prognosis, anti-C1q can be reconsidered for inclusion in classification criteria and in the clinical management of SLE.

## Funding

This work was supported in the form of grants or industrial support from the National Institute of Arthritis and Musculoskeletal and Skin Diseases (NIAMS) and Lupus Foundation of America, and an unrestricted research grant from Human Genome Sciences.

Dr Ana–Maria Orbai is supported by National Institutes of Health (NIH) grant T32 AR048522.

Dr Gunnar Sturfelt and Dr Lennart Truedsson are supported by grants from the Greta and Johan Kock's Foundation, King Gustaf V's 80th Birthday Foundation and the Swedish Rheumatism Association.

Dr Caroline Gordon is supported by Lupus UK.

Dr Paul R. Fortin is supported by a Canada Research Chair on Systemic Autoimmune Rheumatic Diseases.

Dr David A. Isenberg and Dr Anisur Rahman are supported by the National Institute for Health Research University College London Hospitals Biomedical Research Centre.

Dr Sang-Cheol Bae is supported by the Korea Healthcare Technology R & D Project, Ministry for Health and Welfare, Republic of Korea (A120404).

Professor I. Bruce is a National Institute for Health Research (NIHR) Senior Investigator and is supported by Arthritis Research UK, The Manchester Academic Health Science Centre, the NIHR Manchester Musculoskeletal Biomedical Research Unit, The NIHR Manchester Wellcome Trust Clinical Research Facility and the Manchester Biomedical Research Centre. The views expressed in this publication are those of the author(s) and not necessarily those of the NHS, the National Institute for Health Research or the Department of Health.

## Acknowledgment

We thank Mrs. Joyce Kosmas, Johns Hopkins Arthritis Center, for her help with the manuscript submission.

## Conflicts of interest statement

The authors have no conflicts of interest to declare.

## References

- 1 Botto M, Walport MJ. C1q, autoimmunity and apoptosis. *Immunobiology* 2002; 205: 395–406.
- 2 Sturfelt G, Truedsson L. Complement in the immunopathogenesis of rheumatic disease. *Nat Rev Rheumatol* 2012; 8: 458–468.

- 3 Schejbel L, Skattum L, Hagelberg S, *et al.* Molecular basis of hereditary C1q deficiency—revisited: Identification of several novel disease-causing mutations. *Genes Immun* 2011; 12: 626–634.
- 4 Lood C, Gullstrand B, Truedsson L, *et al.* C1q inhibits immune complex-induced interferon-alpha production in plasmacytoid dendritic cells: A novel link between C1q deficiency and systemic lupus erythematosus pathogenesis. *Arthritis Rheum* 2009; 60: 3081–3090.
- 5 Sturfelt G, Johnson U, Sjöholm AG. Sequential studies of complement activation in systemic lupus erythematosus. *Scand J Rheumatol* 1985; 14: 184–196.
- 6 Renal Disease Subcommittee of the American College of Rheumatology Ad Hoc Committee on Systemic Lupus Erythematosus Response Criteria. The American College of Rheumatology response criteria for proliferative and membranous renal disease in systemic lupus erythematosus clinical trials. *Arthritis Rheum* 2006; 54: 421–432.
- 7 Nived O, Hallengren CS, Alm P, Jönsen A, Sturfelt G, Bengtsson A. An observational study of outcome in SLE patients with biopsy-verified glomerulonephritis between 1986 and 2004 in a defined area in Southern Sweden: The clinical utility of the ACR renal response criteria and predictors for renal outcome. *Scand J Rheumatol* 2013; 42: 383–389.
- 8 Mannik M, Wener MH. Deposition of antibodies to the collagen-like region of C1q in renal glomeruli of patients with proliferative lupus glomerulonephritis. *Arthritis Rheum* 1997; 40: 1504–1511.
- 9 Sjöwall C, Olin AI, Skogh T, Wettero J, Morgelin M, Nived O, *et al.* C-reactive protein, immunoglobulin G and complement co-localize in renal immune deposits of proliferative lupus nephritis. *Autoimmunity* 2013; 46(3): 205–214.
- 10 Marto N, Bertolaccini ML, Calabuig E, Hughes GR, Khamashta MA. Anti-C1q antibodies in nephritis: correlation between titres and renal disease activity and positive predictive value in systemic lupus erythematosus. *Ann Rheum Dis* 2005; 64: 444–448.
- 11 Siebert CE, Daha MR, Tseng CM, Coremans IE, van Es LA, Breedveld FC. Predictive value of IgG autoantibodies against C1q for nephritis in systemic lupus erythematosus. *Ann Rheum Dis* 1993; 52: 851–856.
- 12 Trendelenburg M, Lopez-Trascasa M, Potlukova E, *et al.* High prevalence of anti-C1q antibodies in biopsy-proven active lupus nephritis. *Nephrol Dial Transplant* 2006; 21: 3115–3121.
- 13 Fang QY, Yu F, Tan Y, *et al.* Anti-C1q antibodies and IgG subclass distribution in sera from Chinese patients with lupus nephritis. *Nephrol Dial Transplant* 2009; 24: 172–178.
- 14 Moroni G, Radice A, Giammarresi G, *et al.* Are laboratory tests useful for monitoring the activity of lupus nephritis? A 6-year prospective study in a cohort of 228 patients with lupus nephritis. *Ann Rheum Dis* 2009; 68: 234–237.
- 15 Coremans IE, Spronk PE, Bootsma H, *et al.* Changes in antibodies to C1q predict renal relapses in systemic lupus erythematosus. *Am J Kidney Dis* 1995; 26: 595–601.
- 16 Akhter E, Burlingame RW, Seaman AL, Magder L, Petri M. Anti-C1q antibodies have higher correlation with flares of lupus nephritis than other serum markers. *Lupus* 2011; 20: 1267–1274.
- 17 Grootsholten C, Dieker JW, McGrath FD, *et al.* A prospective study of anti-chromatin and anti-C1q autoantibodies in patients with proliferative lupus nephritis treated with cyclophosphamide pulses or azathioprine/methylprednisolone. *Ann Rheum Dis* 2007; 66: 693–696.
- 18 Petri M, Orbai AM, Alarcón GS, *et al.* Derivation and validation of the Systemic Lupus International Collaborating Clinics classification criteria for systemic lupus erythematosus. *Arthritis Rheum* 2012; 64: 2677–2686.
- 19 Martensson U, Sjöholm AG, Sturfelt G, Truedsson L, Laurell AB. Western blot analysis of human IgG reactive with the collagenous portion of C1q: Evidence of distinct binding specificities. *Scand J Immunol* 1992; 35: 735–744.
- 20 Antes U, Heinz HP, Loos M. Evidence for the presence of autoantibodies to the collagen-like portion of C1q in systemic lupus erythematosus. *Arthritis Rheum* 1988; 31: 457–64.
- 21 Wener MH, Uwatoko S, Mannik M. Antibodies to the collagen-like region of C1q in sera of patients with autoimmune rheumatic diseases. *Arthritis Rheum* 1989; 32: 544–551.



- 22 Jonsson G, Sjöholm AG, Truedsson L, Bengtsson AA, Braconier JH, Sturfelt G. Rheumatological manifestations, organ damage and autoimmunity in hereditary C2 deficiency. *Rheumatology (Oxford)* 2007; 46: 1133–1139.
- 23 Petri M, Orbai AM, Alarcón GS, *et al.* Derivation and validation of Systemic Lupus International Collaborating Clinics classification criteria for systemic lupus erythematosus. *Arthritis Rheum* 2012; 64: 2677–2686.
- 24 Horváth L, Cziráj L, Fekete B, *et al.* Levels of antibodies against C1q and 60 kDa family of heat shock proteins in the sera of patients with various autoimmune diseases. *Immunol Lett* 2001; 75: 103–109.
- 25 Mannik M, Merrill CE, Stamps LD, Wener MH. Multiple autoantibodies form the glomerular immune deposits in patients with systemic lupus erythematosus. *J Rheumatol* 2003; 30: 1495–1504.
- 26 Sinico RA, Radice A, Ikehata M, *et al.* Anti-C1q autoantibodies in lupus nephritis: Prevalence and clinical significance. *Ann N Y Acad Sci* 2005; 1050: 193–200.
- 27 Zhang CQ, Ren L, Gao F, Mu FY, You YQ, Liu YH. Anti-C1q antibodies are associated with systemic lupus erythematosus disease activity and lupus nephritis in northeast of China. *Clin Rheumatol* 2011; 30: 967–973.
- 28 Yang XW, Tan Y, Yu F, Zhao MH. Combination of anti-C1q and anti-dsDNA antibodies is associated with higher renal disease activity and predicts renal prognosis of patients with lupus nephritis. *Nephrol Dial Transplant* 2012; 27: 3552–3559.
- 29 Braun A, Sis J, Max R, *et al.* Anti-chromatin and anti-C1q antibodies in systemic lupus erythematosus compared to other systemic autoimmune diseases. *Scand J Rheumatol* 2007; 36: 291–298.
- 30 Trad B, Ben Hassine H, Khalifa M, *et al.* Anti-C1q antibodies and systemic lupus erythematosus in the Tunisian population. *Patho Biol (Paris)* 2013; 61: 113–116.
- 31 Seelen MA, Trouw LA, Daha MR. Diagnostic and prognostic significance of anti-C1q antibodies in systemic lupus erythematosus. *Curr Opin Nephrol Hypertens* 2003; 12: 619–624.
- 32 Heinlen LD, McClain MT, Merrill J, *et al.* Clinical criteria for systemic lupus erythematosus precede diagnosis, and associated autoantibodies are present before clinical symptoms. *Arthritis Rheum* 2007; 56: 2344–2351.
- 33 Siegert CE, Daha MR, Swaak AJ, van der Voort EA, Breedveld FC. The relationship between serum titers of autoantibodies to C1q and age in the general population and in patients with systemic lupus erythematosus. *Clin Immunol Immunopathol* 1993; 67: 204–209.
- 34 Bălănescu E, Tănăsescu C, Bălănescu P, *et al.* Anti C1q antibodies in cutaneous lupus erythematosus. *Rom J Intern Med* 2010; 48: 159–163.